Wizard Gel Extraction

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Quick protocol

1. Run an agarose gel as normal – 1% maximum is best.
2. Excise the gel slice with clean blades and place in eppendorf tube.
3. Weigh the gel slice.
    - A column can accommodate 350mg of gel, if the slice weighs more than this divide to a suitable number of eppendorfs and process separately, pooling the final elution.
4. Add membrane binding solution to the gel slice;
    - Add 1ul/mg of gel slice.
5. Incubate the gel slice in QG buffer at 50-65^{o}C for approx 10 minutes, vortex every couple of minutes.
6. Ensure that the gel has completely dissolved.
7. Place a column in a collection tube.
8. Pipette the dissolved gel slice/buffer mixture into one of the columns.
9. stand at RT for 1minute
10. Centrifuge the column at top speed for 1 minute.
11. Discard the flow through.
12. Add 700µl of membrane wash solution (95% Ethanol added appropriate to kit size) to the column.
13. Centrifuge the column at top speed for 1 minute.
14. Discard the flow through.
15. Add 500µl of membrane wash solution (95% Ethanol added appropriate to kit size) to the column.
16. Centrifuge the column at top speed for 5 minutes.
17. Discard the flow through.

- Take care not to touch the top of the flow through with the base of the column.

18. Centrifuge the column at top speed for 1 minute (this ensures that all the alcohol is removed from the column).

19. Place the column in a fresh tube.

20. Add 50µl nuclease free water directly onto the disc at the base of the column (do not touch the disc).

    - You can add a smaller elution volume to increase the concentration, but the overall yield may be less. Do not use less than 15µl.

21. Stand at RT for 1 minute.

22. Centrifuge at top speed for 1minute.

23. Discard column.

    - To be extra careful transfer the eluted DNA to a clean tube (as the eppendorf lid has been exposed to the inside of the centrifuge)

24. Store eluted DNA at -20 ^{o}C.