# Qiagen Gel Extraction - quick protocol

Prepared by Ms Alex Aitken

1. Run a TAE agarose gel as normal – 1% maximum is best.
2. Excise the gel slice with clean blades and place in eppenorff tube.
3. Add QG buffer to the top of the tube – add at least 1ml; if it does not look like 1ml will fit, cut the gel slice into 2 pieces and put into 2 tubes and process both tubes as individual samples.
4. Incubate the gel slice in QG buffer at 50$^{o}$C for approx 10 minutes, vortex every couple of minutes.
5. Ensure that the gel has completely dissolved.
6. Pipette 750µl into one of the columns.
7. Centrifuge the column for a few secs (typically up to top speed and then stop is sufficient).
8. Discard the flow through and add the remaining sample to the column.
9. Repeat #7.
10. Discard the flow through.
11. Add 750µl of PE buffer (with alcohol added as per instructions on bottle) to the column and leave to stand at RT for 1 minute.
12. Repeat #7.
13. Discard the flow through.
14. Centrifuge the column for 1minute at top speed (this ensures that all the alcohol is removed from the column).
15. Place the column in a fresh tube.
16. Add 25-50µl TE (or use $dH_20$directly onto the disc at the base of the column (do not touch the disc).
17. Stand at RT for 5minutes.
18. Centrifuge at top speed for 1minute.
19. Discard column.
20. Store eluted DNA at -20 $^{o}$C.